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PATENT APPLICATION

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In Re Application of:

JEFFREY KENT FREDENBURGH ET AL.

Application No.: 09/726,032

Filed: November 30, 2000

For: ODOR CONTROL AGENT

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Examiner: D. Ware

Group Art Unit: 1651

November 18, 2004

Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

SUBMISSION OF PRIORITY DOCUMENT

Sir:

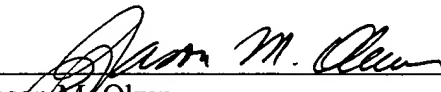
In support of Applicants' claim for priority under 35 U.S.C. § 119, enclosed is
a certified copy of the following Canadian patent application:

CA 2,243,011, filed July 13, 1998.

Applicants request that receipt of this priority document be acknowledged by
the Examiner both in the above-identified application and in Application No. 10/626,537, which
claims priority from the subject application under 35 U.S.C. § 120.

Applicants' undersigned attorney may be reached in our New York office by telephone at (212) 218-2100. All correspondence should continue to be directed to our address given below.

Respectfully submitted,



Jason M. Okun
Attorney for Applicants
Registration No.: 48,512

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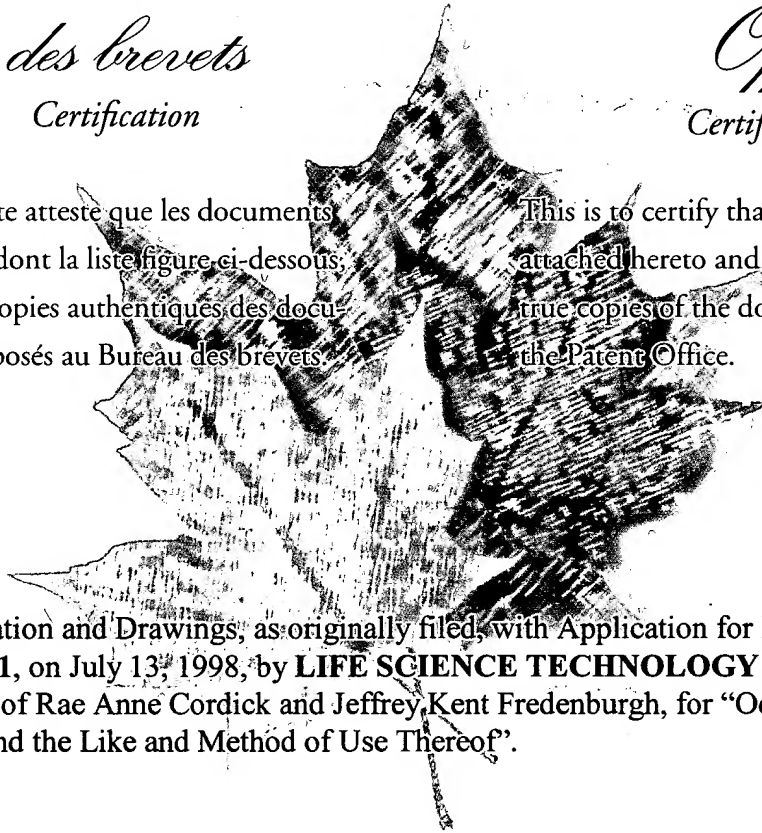
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Specification and Drawings, as originally filed, with Application for Patent Serial No:
2,243,011, on July 13, 1998, by **LIFE SCIENCE TECHNOLOGY GROUP INC.**,
assignee of Rae Anne Cordick and Jeffrey Kent Fredenburgh, for "Odor Control Agent for
Carpet and the Like and Method of Use Thereof".

**CERTIFIED COPY OF
PRIORITY DOCUMENT**


Agent certificateur/Certifying Officer

October 15, 2004

Date

Canada

(CIPO 68)
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ABSTRACT OF THE DISCLOSURE

The present invention provides in one aspect for a method for controlling odor associated with spills of organic material which can cause odors on carpets. The method comprises applying to the carpet a preparation of dormant bacteria, which when activated are effective to control odors. The dormant bacterial preparation is allowed to become associated with the carpet, such that when the carpet is exposed to organic material which can cause odors, the bacteria are capable of becoming active and digesting the organic material.

TITLE: ODOR CONTROL AGENT FOR CARPET AND THE LIKE AND
METHOD OF USE THEREOF

FIELD OF THE INVENTION

5 The present invention is directed to a method of
controlling odor associated with deposits, particularly
spills of organic material on carpet or other fibrous
material and to an odor control agent for use in the
method. The odor control agent can be applied to the
10 carpet or other fibrous material at various stages during
manufacture or use and the effect of the odor control agent
is long lasting.

BACKGROUND OF THE INVENTION

15 Carpet is used extensively in residential and
commercial buildings as it is a relatively inexpensive and
easy to install floor covering material. Carpet offers a
number of desirable qualities including durability,
aesthetics, comfort, safety, warmth and quietness. With
20 modern manufacturing and dyeing techniques, carpeting may
also be provided in almost any color, texture and pattern.
Carpet may be manufactured from diverse types of materials
including natural materials such as wool or cotton or
synthetic materials from various polymers such as
25 polypropylene, polyamide, etc. The majority of carpet,
particularly for residential and commercial use, is
manufactured from synthetic polymer material with
polypropylene and polyamide, most commonly nylon 6 or nylon
66. Irrespective of the material used in the manufacture,
30 the fibers are used in the form of continuous filament
yarns, and in various forms as cut fiber or staple fiber.
One conventional manufacturing process involves inserting
plied yarn into a primary backing of jute or polypropylene
fibers, dyeing the fibers and then applying a carpet
35 backing adhesive such as latex which is adhered to a
secondary backing material.

Many carpet fibers, such as polypropylene and wool, and particularly nylon may be susceptible to staining especially from the many food dyes used in beverages and other foods as well as from other chemicals from many

5 sources. Nylon carpet fibers are often treated with stain blockers such as sulfonated phenol formaldehyde condensate polymer, a sulfonated naphthol formaldehyde condensate polymer, a hydrolyzed vinyl aromatic maleic anhydride polymer or combinations thereof. The stain blockers act to

10 prevent or reduce the ability of organic dyes, particularly acid dye colorants from chemically reacting with and bonding to the nylon. The carpets are also commonly coated with a fluorochemical anti-soiling agent to improve the anti-staining or anti-soiling characteristics of the carpet

15 surface. The fluorochemicals reduce the tendency of soil to adhere to the fiber making the clean-up of any spills or soil on the carpet easier. The fluorochemicals also reduce fiber wettability, making for easy clean up of liquid spills through a simple process of blotting the spill.

20 Examples of such fluorochemicals and other stain resistant chemicals are given, for example, in U.S. Patent numbers 4,680,212 and 4,925,707, the disclosures of which are incorporated herein by reference. The use of the stain blockers and fluorochemicals may not provide complete stain

25 resistance to the carpet, as some materials may still penetrate the nylon fibers or react with the fibers, especially if left in contact with the carpet for extended periods of time. This may be especially true where the carpet is exposed to conditions such as direct sunlight or

30 other UV sources or high traffic areas, as these conditions may cause the effectiveness of the fluorochemical and stain blocker coatings to be diminished.

In addition, especially in residential locations,

35 the possibility of deposits of organic matter such as feces or urine from babies and pets can result in not only soiling of the carpet but also a lingering odor and may, in extreme cases, require the replacement of the carpet. In

the past, various chemical compounds have been proposed to aid in removing odor in a cleaning process. Such chemicals generally act as odor inhibiting agents although U.S. Patent No. 4,946,672 describes the use of biguanidine polymer compositions as odor inhibiting agents. However, even in those cases where the deposit is cleaned up and odor inhibiting agents utilized, the odor from such deposits may remain in the carpet and may become apparent as the effect of the odor masking agents wear off.

10

Deposits of various materials on carpet may also give rise to other concerns. Many of the deposit materials are capable of supporting bacterial growth, especially in the case of feces which contains many bacteria. Some of the bacteria that may grow, as a result of a deposit, may have the potential of causing disease in persons exposed to them, such as mold and mildew. Carpet and other fibrous material are also known to contain a number of naturally occurring bacteria and other organisms. Some of these bacteria may themselves give rise to odor due to incomplete digestion of organic material. There have been attempts to reduce the presence and number of bacteria present in carpet by utilizing various anti-microbial agents such as described in U.S. Patent Nos. 4,110,504 and 5,024,840. These agents are applied to carpet in a manner similar to the way stain blockers are applied to carpet. The use of anti-microbials, while reducing the number of bacteria associated with carpet, may raise other concerns such as the potential that some of the bacteria may become resistant to effects of the anti-microbials.

Many bacterial and fungal genera are known for use in odor control due to their capability for producing enzymes which are capable of breaking down organic material. Such bacteria are particularly useful where the organic material, if allowed to remain, will give rise to malodors. Several such bacterial and fungal genera such as Bacillus, Lactobacillus, Enterobacter, Streptococcus,

Rhizopus, Nitrosomonas, Nitrobacter, Pseudomonas, Alcaligenes and Klebsiella, among others, are known for use in such applications with Bacillus sp. being the most prevalent in use in various applications.

5

For example, European Patent Application No. 732,396 describes the use of Bacillus sp. for odor control of feedstuffs used in farming and JP Patent Application No. 7-031,668 describes their use for odor control of toilets, shoe boxes and pet litter. Other uses of the Bacillus for odor control for baby diapers and wall paper are described in JP Patent Application Nos. 2-121,665 and 3-059,199 respectively. Preparations of active Bacillus in a vegetative form suitable for spraying or otherwise distributing on a deposit, especially of pet urine and feces, on a carpet for controlling odor are presently marketed by The Bramton Company of Dallas, Texas under the trademark OUTRIGHT. The bacterial preparations are used to deodorize a deposit by application directly on the deposit. Once the deposit is deodorized, the bacteria are depleted from the site or disposed of along with the deodorized material. In the event of a new deposit on the carpet, the treatment must be repeated. In all of these circumstances, the Bacillus or other strains of bacteria are used in an active or vegetative state as fully developed bacterial cells capable of immediate growth. It has been thought in the art that the bacteria must be in the active state to be effective, and that dormant or sporulated bacterial forms are ineffective.

30

There thus remains a need for a means for treating carpet and other fibrous material to counteract the effects of deposits and especially for controlling odor associated with the deposits, particularly deposited organic material, where the effects of the odor control are preventative and long lasting.

35

SUMMARY OF THE INVENTION

The present invention provides in one aspect for a method for controlling odor associated with deposits of organic odor causing material on carpets and other fibrous materials. The method comprises applying to the carpet or
5 other fibrous material, a preparation of dormant bacteria, which, when activated, is effective to control odors. The dormant bacterial preparation is allowed to become associated with the carpet or other fibrous material, such that when the carpet or other fibrous material is exposed
10 to organic material which can cause odors, the bacteria are capable of becoming active and digesting the organic material.

In another aspect of the invention there is
15 provided a composition for treating a fabric or fibrous material to provide control of odor associated with deposits of organic odor causing material on the fabric or fibrous material. The composition comprises one or more strains of dormant bacteria, which when activated are
20 effective to control odors.

In yet another aspect of the invention there is provided an aqueous odor controlling bacterial composition for treating carpet or fabric to impart odor control. The
25 composition comprises a stain blocker chemical and an effective amount of odor controlling bacteria.

BRIEF DESCRIPTION OF THE DRAWINGS

Preferred embodiments of the invention are shown in
30 the drawings, wherein:

Figure 1 illustrates scanning electron microscope pictures of carpet fibers containing no innoculum (Fig. 1A) and carpet fibers inoculated with a preferred bacterial spore blend prepared according to Example 1 of the present
35 invention (Fig 1B);

Figure 2 is a graph illustrating the germination and growth of the bacteria spore blend on various organic soils;

Figure 3 is a graph illustrating the germination and growth of the bacterial spore blend in nylon carpet containing various organic soils; and

Figure 4 is a graph illustrating the germination and growth of the bacterial spore blend on carpet containing a combination of fox urine and dog feces.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is directed in one aspect to a method of controlling odor associated with deposits, particularly spills, of organic material which can cause odors on carpet or other fibrous materials. The present invention is also directed to the compositions useful for preparing carpet or other fibrous material to make them capable of controlling odor as well as to the carpet or other fibrous material so prepared. In addition to controlling odor, the compositions may also aid in reducing the staining effects of organic material.

Many bacterial genera are known to produce enzymes which are capable of breaking down organic material. Such bacteria are particularly useful where the organic material, if allowed to remain, will give rise to malodors. Several such bacterial genera such as Bacillus, Lactobacillus, Enterobacter, Streptococcus, Nitrosomonas, Nitrobacter, Pseudomonas, Alcaligenes and Klebsiella amongst others are known for use in such applications, with Bacillus and Lactobacillus sp. being the most prevalent in use in various applications. Strains of bacteria from any of the above noted genera are useful in practicing the present invention. Preferably, the bacterial preparation for use in the present invention is one or more strains of Bacillus or Lactobacillus. More preferably, the strains of bacteria for use in the present invention are selected from Bacillus licheniformis, Bacillus pasteurii, Bacillus laevolacticus and Bacillus amyloliquefaciens. Each of these species have characteristics which make them most effective against particular types of organic materials.

All of these species are capable of enhanced anaerobic and aerobic growth. Bacillus pasteurii is known for superior lipase production, while Bacillus laevolacticus has a very fast germination cycle. Bacillus amyloliquefaciens is high
5 in production of protease enzymes.

The selection of the strains of bacteria for use in the present invention may depend upon many factors. One such factor is the nature of the organic material most
10 commonly expected for the particular application. For example, in a commercial application, the most commonly expected deposits would be soil tracked in from out-of-doors, beverages such as coffee, tea, other food and the like, especially in a restaurant environment, and possibly,
15 inks or toners for printers and other office equipment. Many of these materials are high in fatty components so the bacterial preparation may be enhanced for strains having high activity against such materials. One example of such a bacteria is Bacillus pasteurii known for superior lipase
20 production. In a residential environment, the nature of the deposits may differ with out-of-doors soils. For example, beverages, food and urine and feces from pets and children being most commonly encountered. Depending upon the nature of the deposited material, the preparation may
25 be selected to contain strains having enhanced activity against such materials. Another factor which may affect the nature of the deposit is the geographical location of the installed carpet. This factor would especially relate to the nature of deposits of out-of-doors soil and to the
30 nature of food deposits. Different regions are known to have different soil types and different regions may also have differences in the foods commonly consumed due to cultural and environmental factors. In addition, the temperature of the carpet to be treated will influence the
35 activity of the bacteria. Depending on the strain selected the bacteria will tend to exhibit enhanced activity at higher temperatures. At lower ambient temperatures, more active strains may be desired.

The bacterial preparation will typically comprise one or more strains selected from the genera and species described above. When utilizing a mixture of more than one strain, each of the individual strains may comprise between 3% and 97% of the total of the bacteria present in the preparation. Depending upon the bacteria, these percentages are based on the total cell number or colony forming units or the total mass of the bacterial preparation. For the Bacillus sp. the percentages are based on total cell number. Preferably, each of the strains is present in sufficient numbers to make up 10% to 70% of the total bacteria in the preparation. When mixtures of more than two strains are employed, each of the strains is preferably present in an amount of from 20% to 40% of the total bacteria in the preparation. Particularly preferred preparations for general use in almost all applications are as follows:

20	<u>% of total bacteria</u>		
	<u>Species</u>	<u>Range</u>	<u>Preferred</u>
	<u>Bacillus licheniformis</u>	20-60	40
	<u>Bacillus pasteurii</u>	10-30	20
	<u>Bacillus laevolacticus</u>	10-30	20
25	<u>Bacillus amyloliquefaciens</u>	10-30	20

In a preferred embodiment of the present invention an effective amount of a bacterial composition comprising one or more strains selected from the group consisting of Bacillus licheniformis, Bacillus pasteurii, Bacillus laevolacticus and Bacillus amyloliquefaciens and combinations thereof are provided in a state in which the composition may be applied to a carpet fiber or other fibrous material. The effective amount is a sufficient number of bacteria to provide a relatively uniform coverage of the fiber such that when any portion of the carpet is exposed to a deposit of an odor causing organic material,

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the bacteria will undergo rapid growth and consume the odor causing material. The factors which can affect the number of bacteria to be used relate in most part to the nature of the carpet material. Such factors include the nature of the fiber in terms of the material, e.g. nylon or polypropylene and the like, the characteristics of the yarn in the terms of the denier and number of filaments and the characteristics of the fiber in terms of the number of yarns and the twist. These factors relate to the nature of the carpet in terms of the weight (oz) and height of the pile. All of these factors will affect the amount of exposed surface of the fibers which might be covered by the bacterial preparation. For most applications on carpet, between about 10^6 and 10^8 cells per gram of carpet fiber having a weight between about 20oz and 40oz is most effective with 10^7 cells per gram of carpet fiber being most preferred.

The preparations may be provided as a simple aqueous preparation of a suspension of the Bacillus species in a suitable aqueous carrier, such as in distilled water, tap water, a saline solution or other such aqueous solutions. Preferably, the aqueous composition comprises the odor controlling dormant bacterial strain or strains and an effective amount of a stain blocker. The stain blocker is preferably selected from the group consisting of sulfonated phenol formaldehyde condensate polymer, a sulfonated naphthol formaldehyde condensate polymer, a hydrolyzed vinyl aromatic maleic anhydride polymer or combinations thereof. The aqueous composition may also include one or more fluorochemicals typically utilized for carpet treatment, either on their own or in combination with the stain blocker. Examples of such fluorochemicals include products sold under the trademarks STAINMASTER, STAINMASTER with TEFLON, and ZONYL by DuPont and SCOTCHGARD by 3M.

The selection of the suitable fluorochemicals and stainblocker is well within the knowledge of those of skill in the art. Preferably, the fluorochemicals and stainblockers selected are soluble in water, particularly
5 when the composition is to be used on installed carpet. When utilized during the manufacture of the carpet material, the fluorochemicals and stainblocker may be non-water soluble, provided as a dispersant preparation in which the elevated temperatures during the manufacturing
10 process are used to bind the fluorochemicals and stainblocker to the carpet fiber and affix or attach the bacteria in the process. The use of the stain blocker and/or fluorochemical in the preparation improves the ability of the spores to become associated with the fibers.
15 This provides increased protection of the bacteria from subsequent removal by vacuuming the possible adverse effects of environmental factors. The stain blocker and/or fluorochemical are thought to provide a protective encapsulation of the bacteria to aid in the protection of
20 the bacteria from exposure to potentially harmful conditions such as traffic or the effects of cleaning.

The amount of the stain blocker and/or fluorochemical used in the preparations of the present
25 invention are the amounts typically employed in the carpet and fabric industry and would be well known to those skilled in the art. Ordinarily, depending upon the nature of the stain blocker or fluorochemical and the material being treated and its location, the agents are applied to
30 the material in an amount to result in a treatment rate of about 0.1 wt% to about 20 wt% based upon the weight of the nylon or other fibrous material being treated and the amount of stain blocker and/or fluorochemical. Commonly, the treatment rate will be from about 0.15 wt% to about 10
35 wt%, preferably from about 0.2 wt% to about 4 wt%, more preferably from about 0.25 wt% to about 2 wt%. Most preferably, the stain blocker or fluorochemical are applied to give a treat rate of about 0.25 wt% to 1.0 wt% based

upon the weight of the nylon or other fibrous material being treated.

When used in combination with the stain blocker
5 and/or fluorochemical, the Bacillus species may be provided
as active cells. The term "active cells" is intended to
encompass cells which are in a vegetative form and are
capable of immediate growth when exposed to food sources
usually utilized by the bacteria. The term "dormant cells"
10 is intended to encompass cells which are in a state which
are required to be activated before they can undergo
growth. One example of a dormant cell is a sporulated form
of the bacteria where the spores must undergo activation
and germination before growth of the bacteria can occur.

15

As noted above, due to the protective effects of
the stain blocker and/or fluorochemical, the active
bacteria would be protected from the possible effects of
environmental factors. If the bacteria are provided in an
20 active form, it is thought that they may become dormant
after the application by undergoing sporulation until a
deposit of organic material is encountered. In a preferred
embodiment, the bacteria are provided in an already dormant
or sporulated form. By providing the bacteria in a dormant
25 or sporulated form, the bacteria are further protected from
environmental factors which may prove detrimental to active
bacterial cells. These environmental factors may include
low moisture or humidity, as the carpet or other fibrous
material would generally be kept in a dry state. Other
30 factors may include exposure to heat, chemical agents, UV
radiation from sunlight as well as the exposure to air for
those strains which may be predominantly anaerobic.

The sporulated or dormant strains of bacteria
35 become activated and undergo germination in response to
being exposed to organic material including organic
material which can cause odors. The factors which promote
the activation of the dormant or sporulated bacteria

include the moisture and various organic compounds present in the deposit of organic material. Once activated, the bacteria undergo growth and replication, consuming the organic material in the deposit until the material is
5 consumed. After the material is consumed, the bacteria will then become dormant by undergoing sporulation to await exposure to another deposit of organic material. It is thought that the bacteria will also be somewhat cannibalistic, in that as the bacteria break down after the
10 depletion of the organic material, the degradation products of the break down would be utilized as a food source by other of the bacteria. Once the potential energy source is reduced and the number of bacteria is also reduced, it is thought that the remaining bacteria undergo sporulation to
15 return to a dormant state.

The bacterial preparation may be provided as a concentrate to be diluted with the stain blocker and/or fluorochemical formulation prior to application. If
20 provided as a concentrate, the concentrate may include other agents for improving viability of the bacterial preparation. The concentrate preferably contains between 10 and 20 times the number of cells or spores per ml of the final preparation. To prepare the final preparation 5% to
25 10% by volume of the concentrate is mixed with 90% to 95% by volume of the stain blocker and/or fluorochemical formulation. Thus, each ml of the concentrate is mixed with 10 to 20 ml of the stain blocker and/or fluorochemical formulation to prepare the bacterial preparation for
30 application to carpet and other fibrous material.

When treating carpet, the aqueous odor controlling bacterial composition may be applied to the carpet at any stage during its manufacture. For example, the composition
35 may be utilized to treat the precursor filaments, yarns or fibers prior to their use in the conventional manufacturing process. The filament or yarn may be run through a bath containing the aqueous solution of the bacterial

preparation or the bacterial preparation may be sprayed on the filament. After the treatment, the filaments or yarns are dried and then further processed into carpet in the normal manner. Alternatively, the carpet during the
5 manufacturing process may be immersed, sprayed or otherwise treated with the aqueous composition. The carpet fibers may be sprayed or otherwise treated with the bacterial preparation prior to being inserted into the primary backing. Alternatively, the fibers may also be treated
10 once they have been inserted into the primary backing, either before or after the backing adhesive and secondary backing material have been applied. The composition may also be applied to the finished carpet as a final step prior to drying and rolling. The carpet would be sprayed
15 or otherwise treated with the aqueous bacterial preparation, after which time the carpet would be dried in the usual manner and rolled onto the roll.

Another option would be to apply the composition to
20 an installed carpet. When applying the composition to an installed carpet, it is preferred that the composition be applied thoroughly and evenly throughout the length of the pile, especially reaching down to the base of the pile fiber. This is generally achieved by applying the aqueous
25 bacterial preparation to the carpet and then working the fibers to improve the contact, distribution and penetration of the bacterial preparation. This is most commonly achieved by use of a pile brush operated either by hand or automatically for example, utilizing a cleaning device such
30 as is commonly available commercially. To enhance the penetration of the bacterial preparation, the fibers of the carpet may initially be wetted through an application of a detergent solution. This is most commonly applied where the installed carpet is cleaned using a cleaning machine
35 prior to the application of the bacterial preparation. While the carpet fibers are still moist, the bacterial preparation may be applied and worked into the carpet, utilizing the pile brush. Once the carpet has been so

treated, it is dried, either by allowing it to dry in the air at ambient temperature or through the use of hot air blown through the pile of the carpet to increase the speed of drying of the carpet. Depending upon the state of the carpet or other fibrous material, the composition may be applied in many different ways. The composition may be applied by dipping the material in the composition or by spraying the composition onto the fibrous material. In any of these cases, once the fiber or carpet is treated with the composition, the treated carpet material is allowed to dry by way of applied heat or simply by ambient drying. Alternatively, or in addition to treating the carpet fiber with the aqueous composition, the carpet backing and/or carpet cushion underlayment may also be treated with the bacterial preparation. Once again, the carpet backing and/or carpet cushion underlayment may be treated during the manufacturing process, or prior to its installation. The carpet cushion underlayment may also be similarly treated during the installation of the carpet cushion underlayment.

The following examples illustrate the use of the present invention but are not to be construed as limiting the scope of the present invention.

Example 1

A known weight of carpet was conditioned at 50% humidity at 75°F. After conditioning, the carpet was sprayed with a suspension of a mixture of sporulated forms of Bacillus sp. having the following formulation:

<u>Species</u>	<u>% of total bacteria</u>
<u>Bacillus licheniformis</u>	40
<u>Bacillus pasteurii</u>	20
<u>Bacillus laevolacticus</u>	20
<u>Bacillus amyloliquefaciens</u>	20

The bacterial suspension was prepared in an aqueous solution of 5% ZONYL 7044 fluorochemical in distilled water at a concentration of 10^8 spores per ml. The bacterial suspension was applied to the carpet in an aerosol form to provide a treatment rate of 10^7 spores per gram of carpet. After the application of the bacterial suspension, the carpet was dried at 290°F in an oven in a humidity controlled chamber for 20 minutes. A sample of the carpet fiber treated with the bacterial suspension was compared with a sample of untreated carpet fiber by scanning electron microscopy. The results of this comparison are shown in Figure 1 where Figure 1a illustrates the carpet fiber containing naturally occurring bacteria and other microorganisms adhered to the carpet fiber which was not treated. Figure 1b illustrates a carpet fiber inoculated with the bacterial spore preparation. As can be seen in the micrographs, the treated carpet fiber has a large number of Bacillus spores adhered to the surface of the fiber with very little, if any naturally occurring bacteria or other microorganisms present in the sample.

Example 2

Samples of carpet fiber and plate count broth were examined for oxygen uptake using a standard respirometric study. Oxygen uptake is commonly utilized in those applications where it is not possible to easily measure bacterial growth by other methods. It is known that for aerobic bacteria, oxygen uptake is directly proportional to bacteria count, with the greater the uptake, the higher the corresponding bacteria count would be. The respirometric studies were conducted using a Challenge AER100 respirometer with all samples incubated under controlled temperature conditions. The treatment reactors were 500 ml bottles with CO₂ adsorption trap inserts containing 5 ml of 30% KOH (w/v) with alizarin yellow pH indicator. The sterilized traps were filled with the KOH caustic solution then inserted into the sterilized reactors using aseptic techniques. The CO₂ traps also contained sterilized

medical cotton rolls used as wicks to increase the surface area of the caustic solution. Each reactor was provided with sufficient carpet material to yield 5 grams of carpet fiber. A plate count broth prepared by mixing 17g Difco Plate Count Broth, 0.073g KH_2PO_4 , 0.114g K_2HPO_4 per liter of distilled water and the pH adjusted to 7 was added to the reactor and the reactors autoclaved to sterilize them. The reactors were allowed to cool and 0.5 ml of the bacterial suspension utilized in Example 1 containing 10^8 spores per ml were added to the test reactors. The same volume of distilled water was added to the control reactors. The reactors were capped without the caustic traps and rolled and swirled to ensure that the water and bacterial preparations were mixed well with the organic materials and to permit the carpet to absorb the liquid. The caustic traps were then inserted into the reactors and the reactors hooked up to the respirometer systems. The reactors were incubated in a water temperature bath maintained at 23 °C using an automatic temperature controller. The oxygen uptake by any bacteria growing in the reactors was monitored continuously and reported at 2 hour intervals.

As illustrated in Figure 2, carpet fiber which had not been inoculated with the bacterial spore blend demonstrated only a very slight increase in oxygen uptake after about 24 hours of incubation. The oxygen uptake did not increase above this level up to 60 hours post-inoculation. These results indicate minimal bacterial growth in the control carpet sample. In contrast, the carpet fiber inoculated with the bacterial spore blend showed an increase in oxygen uptake starting 22 to 24 hours after inoculation. This increase in oxygen uptake continued up to the end of the test at 60 hours post-incubation with the oxygen intake increasing in a steady linear fashion with no leveling off of the uptake seen during the 60 hours of the test. These results indicate that the dormant bacteria are capable of germinating to

become active and undergo growth in response to exposure to a suitable food source.

Example 3

5 To confirm that the bacterial spore blend utilized
in the present invention could grow on various organic
soils, plates containing materials representative of common
household or soil causing organic based materials were
inoculated with the bacterial spore blend. The organic
10 based materials utilized were chocolate syrup, tomato
sauce, milk, dog feces and fox urine. The growth on these
soils was compared to a standard plate count broth utilized
for counting colony forming units. The plates were
inoculated with dilutions of the bacterial spore blend to
15 give between about 300 and 400 spores per plate and
incubated at 37°C and 50% humidity. At two days and four
days post inoculation, the colony forming units (CFU) were
counted and the CFU's per ml of the inoculum were
calculated. After two days, the bacterial preparations
20 were growing well on the tomato sauce, chocolate syrup and
dog feces, with growth almost at the level of the standard
plate count broth. A minimal increase in growth on the
autoclaved milk or fox urine was observed after two days,
although there was some growth. After four days, the
25 growth on all five materials was comparable, being only
slightly less than the growth on the plate count broth.
These results indicate that the bacterial spore blend can
grow well on common organic soil, such as chocolate syrup,
tomato sauce, dog feces and fox urine.

30

Example 4

 The bacterial spore blend was tested using
respirometric studies as set out in Example 2 above to
confirm that it could utilize pet waste for growth in
35 carpets. Samples of the carpet fiber were examined for
oxygen uptake using a standard respirometric study
conducted using a Challenge AER100 respirometer with all
samples incubated under controlled temperature conditions.

The treatment reactors were 500 ml bottles. The CO₂ adsorption trap inserts contained 5 ml of 30% KOH (w/v) with alizarin yellow pH indicator. The sterilized traps were filled with the KOH caustic solution then inserted
5 into the sterilized reactors using aseptic techniques. The CO₂ traps also contained sterilized medical cotton rolls used as wicks to increase the surface area of the caustic solution. Each reactor was provided with sufficient carpet material to yield 5 grams of carpet fiber. The organic
10 material (i.e. dog feces, fox urine, plate count broth, etc.) was added to the reactor and the reactors autoclaved to sterilize them. The reactors were allowed to cool and 0.5 ml of the bacterial suspension containing 10⁸ spores per ml were added to the test reactors. The same volume of
15 distilled water was added to the control reactors. The reactors were capped without the caustic traps and rolled and swirled to ensure that the water and bacterial preparations were mixed well with the organic materials and to permit the carpet to absorb the liquid. The caustic
20 traps were then inserted into the reactors and the reactors hooked up to the respirometer systems. The reactors were incubated in a water temperature bath maintained at 23 °C using an automatic temperature controller. The oxygen uptake in the reactors was monitored continuously and
25 reported at 2 hour intervals.

The carpet sample in the control reactor with no inoculum did not have any significant increase in oxygen uptake over the 96 hours of the test. The carpet samples
30 which had been inoculated with the bacterial spore blend started showing an increase in oxygen uptake after 32 hours post-inoculation. This increase in oxygen uptake continued to the end of the test in a linear fashion with no plateauing of the oxygen uptake observed up to 96 hours
35 post-inoculation. This clearly shows that the bacterial spore blend associated with the carpet can become activated and undergo growth when exposed to a common organic spill material.

The compositions and method of the present invention provide for effective odor control for carpet. The use of the bacterial preparations, particularly the sporulated forms of Bacillus, provide for control of odor caused by deposits of organic odor causing material on carpets and other fibrous material. Once the deposit comes into contact with the bacteria, the bacteria germinate if in the dormant form, and commence growing by feeding on the organic material as a food source. As can be observed from the above experiments with the sporulated Bacillus, this bacterial growth commences within about 24 and 48 hours after the bacteria encounter the deposit. In some circumstances, it may be desirable to mask the odor using odor masking agents until the sporulated bacteria can germinate, grow and effectively decompose the odor causing agents. Alternatively, the bacterial preparations may include suitable protease and lipase enzymes to commence the digestion of the odor causing material until the bacteria commence their growth stage and can take over the digestion of the odor causing material. As a further alternative, introducing suitable molecular sieves that can quickly bind the offensive odor within its' pores, allowing time for the sporulated bacteria to decompose the odor causing material.

It has surprisingly been found that the odor control agent applied to the carpet as described above remains effective for extended periods of time even with carpet exposed to high traffic and repeated vacuuming. The exact mechanism for this is not completely understood, but it is suspected that the dormant bacteria become so tightly associated with the carpet fibers that they are not easily removed when exposed to traffic or vacuuming. The use of the stain blocker and/or fluorochemical in the aqueous solution used in the application of the agent to the carpet is suspected to increase the association of the dormant bacteria with the carpet fibers, and hence increase the

effective life of the treatment. It is suspected that the treatment may also remain after wet cleaning of the carpet. However, it is suggested that the carpet be treated with the odor controlling bacterial preparation on a routine
5 basis such as after each wet cleaning. This can be easily accomplished after cleaning with the preparation applied to the carpet either when still wet from the cleaning or after the carpet has dried. Preferably, the preparation is applied to the carpet while still wet, worked into the
10 carpet with a pile brush and the carpet is allowed to dry naturally.

In addition to providing for removal of potentially odor causing organic material associated with deposits on
15 carpet and other fibrous material, the use of the bacterial preparations of the present invention provides other benefits. Based upon the observations from the electron micrographs, it is expected that the presence of the bacterial preparation in association with the carpet fiber
20 and other fibrous material may result in a reduction in the presence of other bacteria and organisms which are naturally found on installed carpet and other fibrous material, both in number and population. It has also been found that the bacterial preparation associated with the
25 carpet fiber or other fibrous material enhances the anti-stain characteristics of the carpet. Many of the stain causing materials are organic in nature and it has been found that the bacteria can utilize such organic materials as a food source. As the stain causing material is
30 consumed by the bacteria, the staining properties of the compounds are reduced.

The method and compositions of the present invention are especially suitable for use with carpet as
35 described in the specific examples set out above. These methods and compositions are also suitable for use with other fibrous material which may be susceptible to the effects of deposits of organic material. Examples of such

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other fibrous materials include rugs, upholstery fabrics, automotive fabrics, bedding, clothing, etc. Suitable binders may be determined to improve the longevity and efficacy to address wash and wearing performance. Other
5 applications may include hard surfaces, such as ceramics, tile, walls, wood, etc.

Although various preferred embodiments of the present invention have been described herein in detail, it
10 will be appreciated by those skilled in the art, that variations may be made thereto without departing from the spirit of the invention or the scope of the appended claims.

THE EMBODIMENTS OF THE INVENTION IN WHICH AN EXCLUSIVE
PROPERTY OR PRIVILEGE IS CLAIMED ARE DEFINED AS FOLLOWS:

1. A method for controlling odor associated with
5 deposits of organic material which can cause odors on
carpets or other fibrous material, the method comprising
applying to the carpet or other fibrous material or to the
fibers used in the manufacture of the carpet or other
10 fibrous material, a preparation of dormant bacteria, which
when activated are effective to control odors, the dormant
bacterial preparation being allowed to become associated
with the carpet or other fibrous material such that when
the carpet or other fibrous material is exposed to organic
15 material which can cause odors, the bacteria are capable of
becoming active and digesting the organic material.
2. A method as claimed in claim 1 wherein the dormant
bacteria are sporulated forms of one or more strains
selected from the bacterial genera Bacillus.
- 20 3. A method as claimed in claim 1 wherein the dormant
bacteria are sporulated forms of one or more strains
selected from the group of bacterial species consisting
essentially of Bacillus licheniformis, Bacillus pasteurii,
25 Bacillus laevolacticus and Bacillus amyloliquefaciens.
4. A method as claimed in claim 3 wherein the dormant
bacteria are applied to the carpet at a concentration of
between about 10^6 and about 10^8 cells per gram of carpet
30 fiber.
5. A method as claimed in claim 4 wherein the dormant
bacteria are applied to the carpet at a concentration of
about 10^7 cells per gram of carpet fiber.

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6. A method as claimed in claim 3 wherein the dormant bacterial preparation comprises:

5	<u>Species</u>	<u>% of total bacteria</u>	
		<u>Range</u>	<u>Preferred</u>
	<u>Bacillus licheniformis</u>	20-60	40
	<u>Bacillus pasteurii</u>	10-30	20
	<u>Bacillus laevolacticus</u>	10-30	20
	<u>Bacillus amyloliquefaciens</u>	10-30	20

10

7. A method as claimed in claim 3 wherein the bacterial preparation includes one or more stain-blocking chemicals.

15

8. A method as claimed in claim 7 wherein the one or more stain-blocking chemicals are selected from the group consisting of sulfonated phenol formaldehyde condensate polymer, sulfonated naphthol formaldehyde condensate polymer, and hydrolyzed vinyl aromatic maleic anhydride polymer.

20

9. A method as claimed in claim 8 wherein the preparation contains an amount of the stain blocker to result in a treat rate of the carpet of about 0.1 wt% to about 20 wt% based upon the weight of the carpet fiber.

25

10. A method as claimed in claim 9 wherein the treat rate is from about 0.25 wt% to about 20 wt%.

30

11. A method as claimed in claim 8 wherein the bacterial preparation further includes one or more anti-soil fluorochemicals.

12. An aqueous odor controlling bacterial composition for treating carpet or other fibrous material to impart odor control to the carpet or other fibrous material, the

35

composition comprising one or more stain-blocker chemicals and an effective amount of odor controlling bacteria.

13. An aqueous odor controlling bacterial composition as claimed in claim 12 wherein the bacteria are one or more strains selected from the group of bacterial genera consisting of Bacillus, Enterobacter, Streptococcus, Nitrosomonas, Nitrobacter, Pseudomonas, Alcaligenes and Klebsiella.

10

14. An aqueous odor controlling bacterial composition as claimed in claim 13 wherein the bacteria are one or more strains selected from the group of bacterial species consisting essentially of Bacillus licheniformis, Bacillus pasteurii, Bacillus laevolacticus and Bacillus amyloliquefaciens.

15. An aqueous odor controlling bacterial composition as claimed in claim 14 wherein the bacteria are applied to the carpet or other fibrous material at a concentration of between about 10^6 and about 10^8 cells per gram of carpet fiber.

16. An aqueous odor controlling bacterial composition as claimed in claim 15 wherein the dormant bacteria are applied to the carpet or other fibrous material at a concentration of about 10^7 cells per gram of carpet fiber.

17. An aqueous odor controlling bacterial composition as claimed in claim 14 wherein the bacterial preparation comprises:

	<u>Species</u>	<u>% of total bacteria</u>	
		<u>Range</u>	<u>Preferred</u>
	<u>Bacillus licheniformis</u>	20-60	40
35	<u>Bacillus pasteurii</u>	10-30	20
	<u>Bacillus laevolacticus</u>	10-30	20
	<u>Bacillus amyloliquefaciens</u>	10-30	20

18. An aqueous odor controlling bacterial composition as claimed in claim 14 wherein the one or more stain-blocking chemicals are selected from the group consisting of sulfonated phenol formaldehyde condensate polymer, sulfonated naphthol formaldehyde condensate polymer, and hydrolyzed vinyl aromatic maleic anhydride polymer.

19. An aqueous odor controlling bacterial composition as claimed in claim 18 wherein the preparation contains an amount of the stain blocker to result in a treat rate of the carpet of about 0.1 wt% to about 20 wt% based upon the weight of the carpet fiber.

20. An aqueous odor controlling bacterial composition as claimed in claim 19 wherein the treat rate is from about 0.25 wt% to about 20 wt%.

21. An aqueous odor controlling bacterial composition as claimed in claim 20 wherein the bacterial composition further includes one or more anti-soil fluorochemicals.

22. A carpet capable of controlling odor associated with deposits of organic material which can cause odors on the carpet, the carpet comprising fibers tufted through a primary backing, the fibers having associated therewith a preparation of dormant bacteria, which when activated are effective to control odors, such that when the carpet is exposed to organic material which can cause odors, the bacteria are capable of becoming active and digesting the organic material.

23. A carpet as claimed in claim 23 wherein the bacteria are one or more strains selected from the group of bacterial genera Bacillus.

24. A carpet as claimed in claim 22 wherein the bacteria are one or more strains selected from the group of bacterial species consisting essentially of Bacillus

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licheniformis, Bacillus pasteurii, Bacillus laevolacticus
and Bacillus amyloliquefaciens.

25. A carpet as claimed in claim 24 wherein the dormant
5 bacteria are applied to the carpet at a concentration of
between about 10^6 and about 10^8 cells per gram of carpet
fiber.

26. A carpet as claimed in claim 25 wherein the dormant
10 bacteria are applied to the carpet at a concentration of
about 10^7 cells per gram of carpet fiber.

27. A carpet as claimed in claim 24 wherein the dormant
bacterial preparation comprises:

15	<u>% of total bacteria</u>		
	<u>Species</u>	<u>Range</u>	<u>Preferred</u>
	<u>Bacillus licheniformis</u>	20-60	40
	<u>Bacillus pasteurii</u>	10-30	20
	<u>Bacillus laevolacticus</u>	10-30	20
20	<u>Bacillus amyloliquefaciens</u>	10-30	20

28. A carpet as claimed in claim 24 wherein the carpet
has also been treated with one or more stain-blocking
chemicals.

25
29. A carpet as claimed in claim 28 wherein the one or
more stain-blocking chemicals are selected from the group
consisting of sulfonated phenol formaldehyde condensate
polymer, sulfonated naphthol formaldehyde condensate
30 polymer, and hydrolyzed vinyl aromatic maleic anhydride
polymer.

30. A carpet as claimed in claim 29 wherein the
preparation contains an amount of the stain blocker to
35 result in a treat rate of the carpet of about 0.1 wt% to
about 20 wt% based upon the weight of the carpet fiber.

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31. A carpet as claimed in claim 30 wherein the treat rate is from about 0.25 wt% to about 20 wt%.

32. A carpet as claimed in claim 29 wherein the carpet
5 has also been treated with one or more anti-soil
fluorochemicals.



FIG 1A

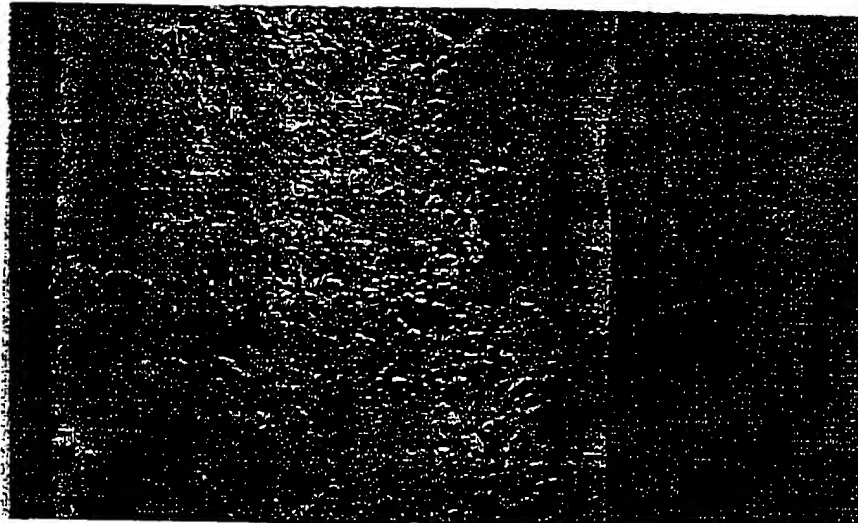


FIG 1B

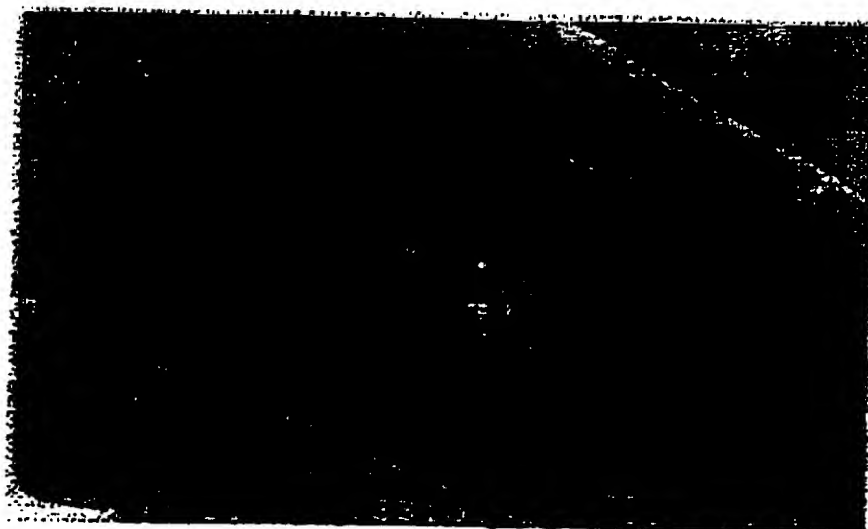


FIG 1A

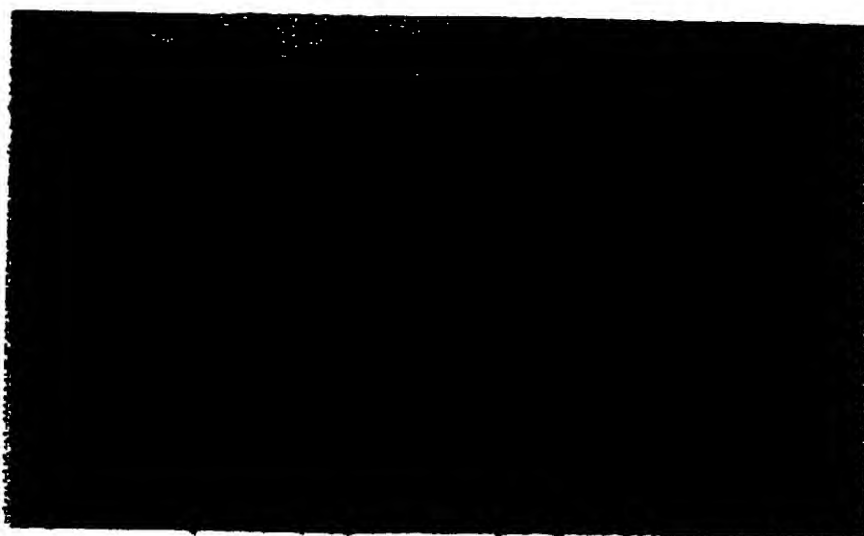


FIG 1B

FIG 2

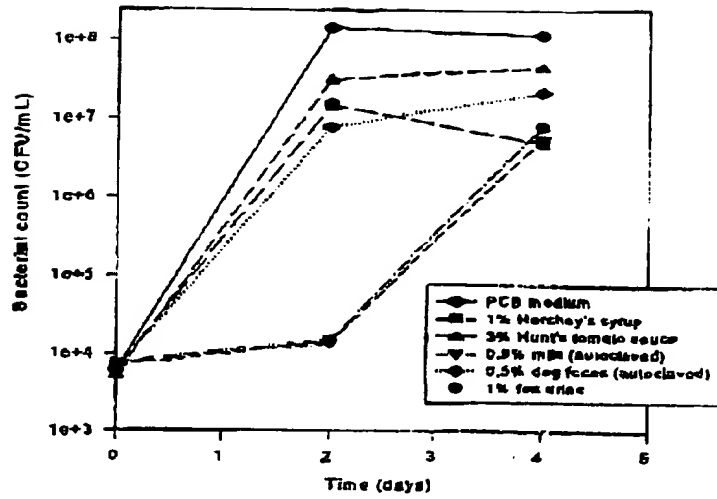


FIG 3

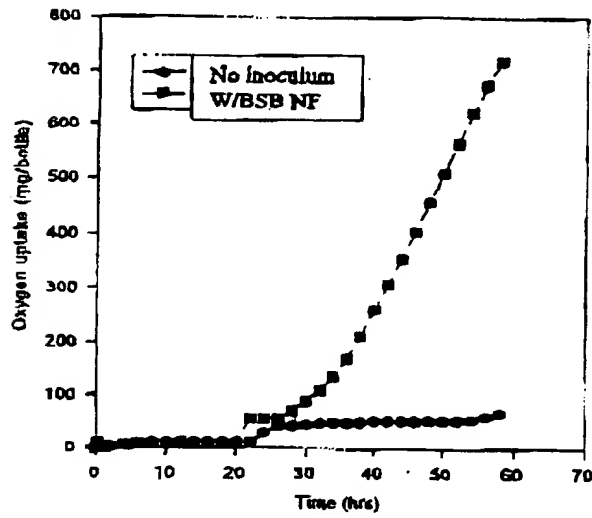


FIG 4

